

Synthesis and evaluation of anti viral, anti tubercular and anticancer activities of some novel thioureas derived from 4-aminobenzohydrazidehydrazones

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ABSTRACT: A class of unique 1-[4-[[2-[(4-substituted phenyl)methylene]hydrazino]carbonyl]p henyl] (By condensing 4-aminobenzoic acid hydrozide with 4-fluorobenzaldehyde or 4-(trifluoromethyl)benzaldehyde, substituted aryl isothiocyanates were added to 4-amino-N'-[(4-substituted phenyl) methylene] benzohydrazide, resulting in -3-substituted thiourea derivatives. We used HeLa, Vero, or HEL cell cultures to test all of the produced compounds in vitro against HIV-1 (IIIB) and HIV-2 (ROD) strains in MT-4 cells, along with additional chosen viruses including HSV-1, HSV-2, Coxsackie virus B4, Sindbis virus, human cytomegalovirus, and varicella-zoster virus. They also tested it for antimycobacterial activity against Mycobacterium tuberculosis H37Rv. The synthetic compounds were tested for anticancer activity and cytotoxicity using A549 and L929 cell lines.

KEY WORDS: Hydrazones, Thioureas, Antiviral activity, Anticancer activity, Mycobacterium tuberculosis H37Rv

INTRODUCTION

There have been reports of thiacetazone, a tuberculostatic drug, which has a thiosemicarbazone structure (1). A well-known inhibitor of Mycobacterium tuberculosis is thiocarlide, which is N,NL-bis[p-(isoamyloxy)phenyl]-thiourea (2). Thiocarlide structural analogues, N-D-aldopentofuranosyl-NL-[p-(isoamyloxy)phenyl] thiourea derivatives, have been shown to be more effective than thiocarlide itself, according to recent reports (3). As seen in Figure 1, methisazone was one of the first antiviral compounds to be utilized in an inclinational fashion. This medicine is effective in preventing the spread of many viral infections. Several 4-aminobenzoic acid-substituted benzalhydrazones have shown antitubercular properties (5). The antioxidant activity of several thiourea derivatives derived from isonicotinoyl hydrazone has been recently reported by Sriram and colleagues.

(6). Some hydrazide-hydrazones and thioureas show antitumor (7,8) and antitubercular (9–11) effects. A number of thiourea derivatives have also been shown to inhibit the activity of influenza virus nuclease, coxsackie B4 virus, and thymidine kinase positive varicella-zoster virus (TK+ VZV, OKA strain) (12,13).

In continuation of our earlier work on 4-aminobenzoic acid hydrazones (14) and various thiourea derivatives (15,16), we have synthesized a series of novel thioureas incorporating hydrazide-hydrazone and disubstituted thiourea moieties into a single structure. These were derived from 4-amino-N'-[[4-fluoro/4-(trifluoromethyl)phenyl]methylene] benzohydrazide, and their antitubercular, antiviral, and anticancer efficacy were assessed. We used HeLa, Vero, or human embryonic lung (HEL) cells to screen all of the synthesized compounds in vitro against HIV-1 (IIIB) and HIV-2 (ROD) strains in MT-4 cells, as well as other selected viruses like HSV-1, HSV-2, Coxsackie B4 virus, Sindbis virus, cytomegalovirus (CMV), and varicella-zoster virus (VZV).

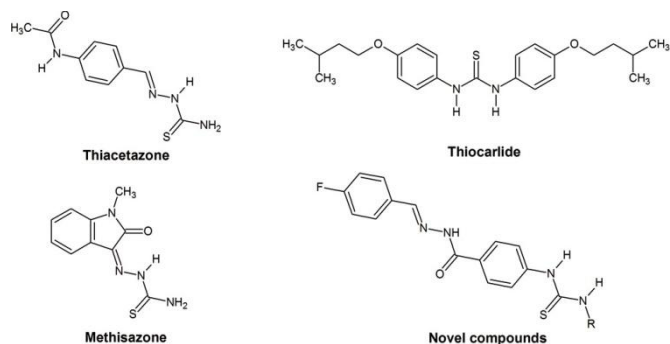


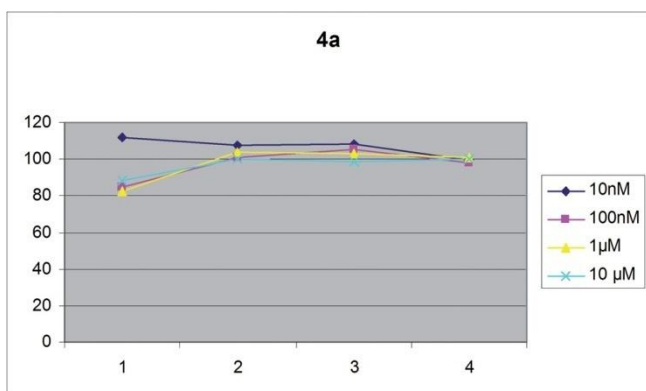
FIGURE1. Some thioureas with antimycobacterial or antiviral activity.

In vitro antitubercular activity of novel compounds against *Mycobacterium tuberculosis* H37Rv was evaluated at TAACF. Anticancer potential of the synthesized compounds was determined using the A 549 and L 929 cell lines.

RESULTS AND DISCUSSION

Chemistry

4-Aminobenzoic acid hydrazide **1** was prepared by the reaction of ethyl 4-aminobenzoate with hydrazine hydrate. 4-Amino-*N'*-[[4-fluoro/4-(trifluoromethyl)phenyl]methylene]benzohydrazide **2-3** (cf. Experimental Section) were synthesized by condensation of **1** with 4-fluoro (17) / 4-(trifluoromethyl)benzaldehyde. 1-[4-[[2-[(4-Substitutedphenyl)methylene]hydrazino]carbonyl]phenyl]-3-substituted thioureas **4a-g** and **5a-f** were synthesized by the reaction of 4-amino-*N'*-[[4-fluoro/4-(trifluoromethyl)phenyl]methylene]benzohydrazide



in a mixture of dry acetonitrile and substituted phenyl isothiocyanates produces yields ranging from 42% to 68% per scheme. It was found that certain dry solvents or mixes might be used to carry out the thiourea reaction (18-21). The current investigation demonstrated that dry acetonitrile was beneficial. Tables 1 and 2 provide the physical and spectral data of thioureas 4a-g and 5a-f, respectively.

They were chosen as starting compounds to design several new thioureas, including 4-amino-*N'*-[(4-fluorophenyl)methylene]benzohydrazide, which was originally synthesized in the present study, and 4-amino-*N'*-[[4-(trifluoromethyl)phenyl]methylene]benzohydrazide, which was reported to have inhibitory potency against *Mycobacterium tuberculosis* H37Rv at 12.5 µg/ml (14). Single signals corresponding to azomethine proton resonances at 8.38-8.51 ppm were seen in the ¹H-NMR spectra of 4a-g and 5a-f (22). The predicted thiourea structures are supported by observations in ¹H-NMR spectra, such as resonances at 8.44-10.03 and 9.84-10.24 ppm corresponding to thiourea R-NH-CS- and -CS-NH-Ar functions (15), respectively. It is worth noting that there are no resonances related to NH₂ function. The predicted values were likewise recorded for the other chemical changes.

With a bias of less than 8 mmu between the experimental and estimated m/z values of either fragment ions or

molecules, high resolution mass spectra (HRMS) verified the molecular weights and empirical formula of compounds 4a-g and 5a-f (Table 2). While compound 4c showed molecular ion peaks when utilizing electron impact (EI) as its ionization mechanism, the other compounds did not. The rapid atomic bombardment (FAB) method, which yields precise MH^+ peaks rather than M^+ in a 3-nitrobenzylalcohol matrix, was used to examine these compounds. The predicted structure was further confirmed by the fragmentation pattern of the sample molecule 4c, as shown in Scheme 2. The first breakage occurred during thiourea moiety cleavage, resulting in the isothiocyanate fragment at m/z 299.0529 due to observed benzyl loss. Additionally, hydrazide-hydrazone characteristic fragmentations were noted. The base peak at m/z 120.0444 was caused by the main fragmentation product, which was identified as a 4-aminophenyl carbo-nyl cation.

Antiviral activity

Compounds 4a-g and 5a-f were tested for antiviral activity and cytotoxicity in various viral test systems (Tables 3-5), according to previously published procedures (23-27). The following viruses and host cells were used for the evaluation :

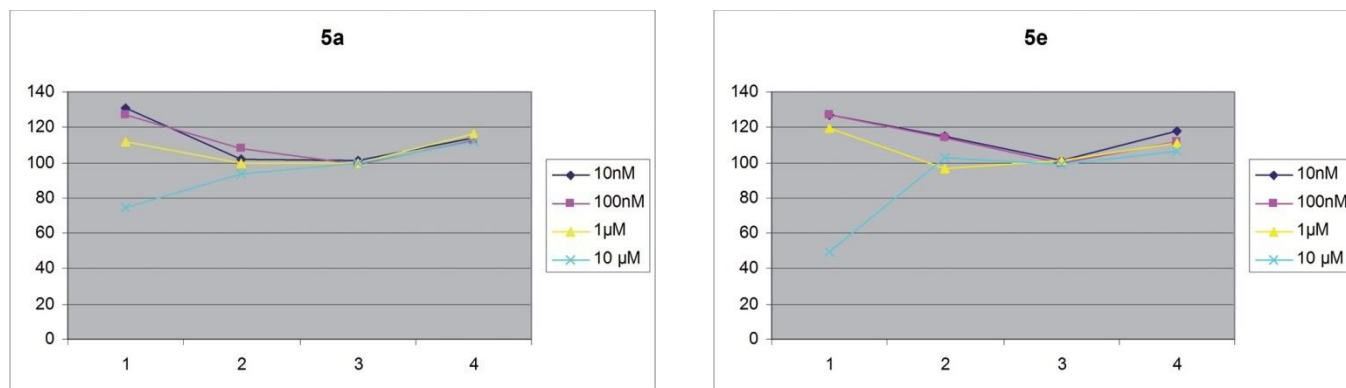
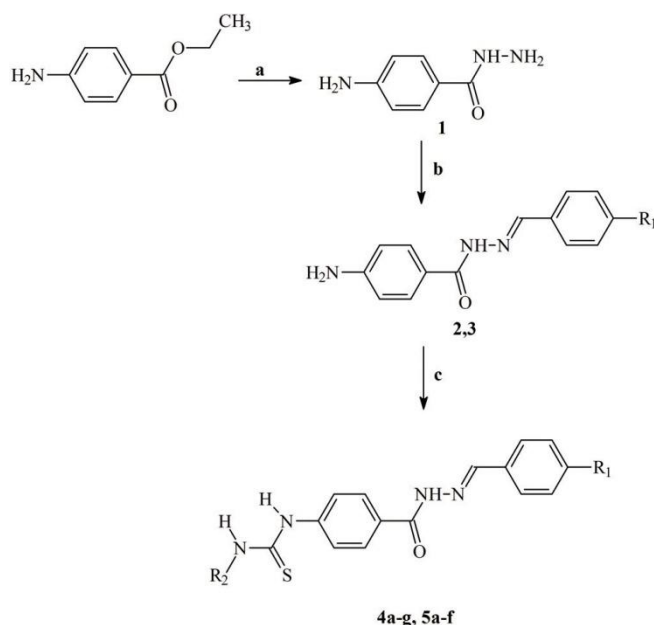


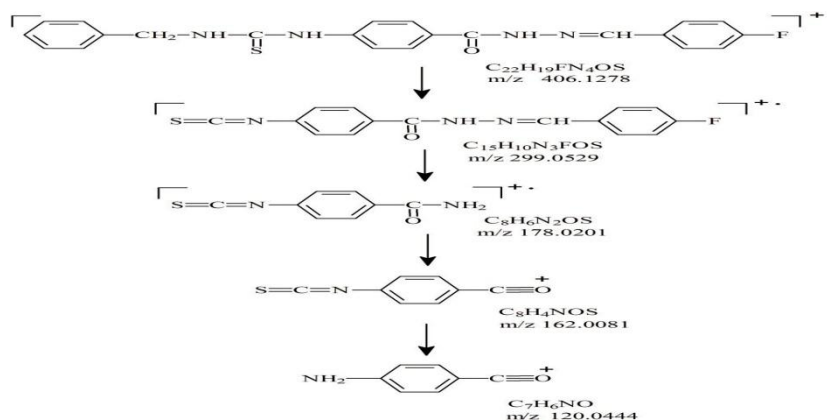
FIGURE 2. Cytotoxic effects of the compounds 4a, 5a, 5e at four different concentrations (10nM, 100nM, 1µM, 10µM).



SCHEME 1. Synthetic route to compounds 2, 3, 4a-g and 5a-f. Reagents and conditions: (a) H_2N-NH_2 , $H_2O/EtOH$, reflux; (b) $R_1-C_6H_4-CH=O/EtOH$, reflux; (c) $R_2-C_6H_4-NCS$ /dry acetonitrile, reflux.

- (a) Vero cell cultures : Parainfluenza-3 virus, Reovirus-1, Sind-bis virus, Punto Toro virus and Coxsackie B4 virus.
(b) HeLa cell cultures : Vesicular stomatitis virus (VSV), Cox-sackie B4 virus and respiratory syncytial virus.

TABLE1 .Physicalpropertiesandelementalanalysisdataof4a-gand5a-f.



Compd	R1	R2	Formula	M.W.	Color M.p(°C)	Yield*(%)	ElementalAn alysis (Calculated /Found)			
							C	H	N	S
4a	-F	-C6H5	C ₂₁ H ₁₇ FN ₄ OS.½ H ₂ O	401.456	White 238	42	62.83 62.57	4.52 4.08	13.96 14.60	7.98 8.30
4b	-F	-C ₆ H ₄ - OCH ₃	C ₂₂ H ₁₉ FN ₄ O ₂ S	422.475	White 231-4	55	62.54 61.94	4.53 4.17	13.26 13.83	7.59 7.75
4c	-F	- CH ₂ C ₆ H 5	C ₂₂ H ₁₉ FN ₄ OS	406.476	White 242	58	65.01 64.28	4.71 4.52	13.78 13.81	7.89 8.77
4d	-F	-C ₆ H ₄ -Br	C ₂₁ H ₁₆ BrFN ₄ OS	471.345	Whitish cream 240	59	53.51 53.09	3.42 3.30	11.89 11.75	6.80 7.72
4e	-F	-C ₆ H ₄ -Cl	C ₂₁ H ₁₆ ClFN ₄ OS	426.894	White 240	67	59.08 58.88	3.78 3.66	13.12 13.14	7.51 8.24
4f	-F	-C ₆ H ₄ -F	C ₂₁ H ₁₆ F ₂ N ₄ OS. ½H ₂ O	419.448	Whitish cream 244	68	60.13 60.26	4.08 3.96	13.35 14.39	7.64 7.33
4g	-F	-C ₆ H ₄ - CH ₃	C ₂₂ H ₁₉ FN ₄ OS.½ H ₂ O	415.486	White 235	45	63.60 63.99	4.85 4.50	13.48 13.94	7.72 8.11
5a	-CF ₃	-C ₆ H ₅	C ₂₂ H ₁₇ F ₃ N ₄ OS. ½H ₂ O	451.464	White 265-8	46	58.53 58.50	4.02 3.65	12.41 12.56	7.10 7.21
5b	-CF ₃	C ₆ H ₄ - OCH ₃	C ₂₃ H ₁₉ F ₃ N ₄ O ₂ S. ½H ₂ O	481.490	White	47	57.37	4.19	11.64	6.66

<i>5c</i>	-CF ₃ -C ₆ H ₄ -Br	$C_{22}H_{16}BrF_3N_4O$ S	521.353	White	59	57.43	3.63	12.28	6.03
<i>5d</i>	-CF ₃ -C ₆ H ₄ -Cl	$C_{22}H_{16}ClF_3N_4O$ S	476.902	Whitish cream	58	50.72	3.13	10.86	6.63
<i>5e</i>	-CF ₃ -C ₆ H ₄ -F	$C_{22}H_{16}F_4N_4OS$ $\frac{1}{2}H_2O$	469.455	White	57	54.96	3.36	11.71	7.27
<i>5f</i>	-CF ₃ -C ₆ H ₄ - CH ₃	$C_{23}H_{19}F_3N_4OS$ H ₂ O	419.514	White	54	55.94	3.43	12.66	6.38
						245	56.39	3.78	12.06
						245	56.39	3.78	12.06

SCHEME2.HR-EI massspectral fragmentation of 4c.

TABLE2. IR, ¹H-NMR and HR mass spectral data of 4a-g and 5a-f.

Compd	IRν(cm ⁻¹) NH, C=O, C=S	¹ H-NMR(DMSO-d ₆ , ppm)	HR-MS(m/z)	
			Calculated	Found
4a	3321,1655,1238	7.12-7.90(m,13H,Ar-H);8.44(s,1H,CH=N);9.96-10.07(d,1H,NH-CS-); 10.22(b,1H,NH-CS-);11.78(d,1H,CO-NH).	393.1180	(FAB) 393.1202(MH ⁺)
4b	3317,3236,1650,1238	3.74(t,3H,O-CH ₃);6.90-7.90(m,12H,Ar-H);8.44(s,1H,CH=N);9.81,9.84	423.1286	(FAB) 423.1328(MH ⁺)
4c	3283,1651,1234	4.75(d,2H,N-CH ₂);7.24-7.87(m,13H,Ar-H);8.38-8.44(d,2H,CH=N)and	406.1264	(EI) 406.1278(M ⁺)
4d	3294,3232,1654,1242	7.25-7.87(m,12H,Ar-H);8.43(s,1H,CH=N);10.01-10.07(d,2H,NH-CS- NH); 11.77 (s, 1H, CO-NH).	471.0285	(FAB) 471.0301(MH ⁺) 473.0308(MH ⁺ +2)
4e	3320,3236,1659,1245	7.24-7.87(m,12H,Ar-H);8.44(s,1H,CH=N);10.02-10.07(d,2H,NH-CS- NH); 11.76 (s, 1H, CO-NH).	427.0790	(FAB) 427.0827(MH ⁺) 429.0705(MH ⁺ +2)
4f	3217,1643,1238	7.14-7.90(m,12H,Ar-H);8.44(s,1H,CH=N);9.91,10.05(2s,1H,NH-CS-); 10.21(s,1H,NH-CS-);11.76-11.78(d,1H,CO-NH).	411.1086	(FAB) 411.1108(MH ⁺)
4g	3321,3236,1655,1238	2.27(s,3H,C ₆ H ₄ CH ₃);7.13-7.90(m,12H,Ar-H);8.44(s,1H,CH=N);9.87-	407.1336	(FAB) 407.1324(MH ⁺)
5a	3321,3236,1655,1238	7.12-7.94(m,13H,Ar-H);8.51(s,1H,CH=N);9.97-10.02(d,1H,NH-CS); 10.24(b1H,NH-CS-);11.95(s,1H,CO-NH).	443.1148	(FAB) 443.1132(MH ⁺)
5b	3301,3240,1651,1249	3.83(t,3H,O-CH ₃);6.90-7.92(m,12H,Ar-H);8.51(s,1H,CH=N);9.79-9.86	473.1254	(FAB) 473.1258(MH ⁺)
5c	3309,3232,1655,1257	7.42-7.94(m,12H,Ar-H);8.51(s,1H,CH=N);10.03(s,1H,NH-CS);10.10(s, 1H,CS-NH);11.95(s,1H,CO-NH).	521.0253	(FAB) 521.0250(MH ⁺) 523.0242(MH ⁺ +2)
5d	3309,1655,1172	7.38-7.94(m,12H,Ar-H);8.51(s,1H,CH=N);10.03-10.09(d,2H,NH-CS- NH); 11.96 (s, 1H, CO-NH).	477.0758	(FAB) 477.0743(MH ⁺) 479.0733(MH ⁺ +2)
5e	3317,3202,1651,1172	7.15-7.92(m,12H,Ar-H);8.51(s,1H,CH=N);9.92-10.02(2s,1H,NH-CS); 10.24(s,1H,CS-NH);11.97(s,1H,CO-NH).	461.1054	(FAB) 461.1049(MH ⁺)
5f	3317,3236,1651,1165	2.32(s,3H,C ₆ H ₄ CH ₃);7.13-7.92(m,12H,Ar-H);8.51(s,1H,CH=N);9.91-	457.1304	(FAB) 457.1335(MH ⁺)

10.24(d,b,2H,NH-CS-NH);11.95(s,1H,CO-NH).

TABLE3. Cytotoxicity and antiviral activity of compounds in Hel, HeLa and Vero cell cultures

Compound	HEL cell cultures						HELA cell cultures				Vero cell cultures					
	Min.	Min. inhibitory conc. ^b (μg/ml)					Min.	Min. inhibitory conc. ^b (μg/ml)			Min.	Min. inhibitory conc. ^b (μg/ml)				
cytoto xic con c.a(μ g/ml)	Her pessi mple x viri s-1 (KO S)	Her pessi mple x viri s-2 (G)	Vacci nia viri s	Vesicu lar attis vir us	Herp essi mple x viri s TKK OSA CVr	cytoto xic a conc. (μg/m l)	Vesicul ar stoma titis viri s	Coxsac kieB4 virus	Respirato rysincyt ial virus	cytoto xic a conc. (μg/m l)	Para influenz a-3 virus	Re ovirus- 1	Sindb viri s	Coxsac kieB4 virus	Pun taT oro viri us	
4a	8	>1.6	>1.6	>1.6	>1.6	>1.6	40	>8	>8	>8	40	>8	>8	>8	>8	
4b	8	>1.6	>1.6	>1.6	>1.6	>1.6	40	>8	>8	>8	40	>8	>8	>8	>8	
4c	8	>1.6	>1.6	>1.6	>1.6	>1.6	8	>1.6	>1.6	>1.6	8	>1.6	>1.6	>1.6	>1.6	
4d	8	>1.6	>1.6	>1.6	>1.6	>1.6	200	>40	>40	>40	40	>8	>8	>8	>8	
4e	8	>1.6	>1.6	>1.6	>1.6	>1.6	40	>8	>8	>8	40	>8	>8	>8	>8	
4f	8	>1.6	>1.6	>1.6	>1.6	>1.6	40	>8	>8	>8	40	>8	>8	>8	>8	
4g	8	>1.6	>1.6	>1.6	>1.6	>1.6	40	>8	>8	>8	40	>8	>8	>8	>8	
5a	8	>1.6	>1.6	>1.6	>1.6	>1.6	40	>8	>8	>8	40	>8	>8	>8	>8	
5b	8	>1.6	>1.6	>1.6	>1.6	>1.6	200	>40	>40	>40	40	>8	>8	>8	>8	
5c	8	>1.6	>1.6	>1.6	>1.6	>1.6	200	>40	>40	>40	40	>8	>8	>8	>8	
5d	8	>1.6	>1.6	>1.6	>1.6	>1.6	40	>8	>8	>8	40	>8	>8	>8	>8	
5e	8	>1.6	>1.6	>1.6	>1.6	>1.6	200	>40	>40	>40	40	>8	>8	>8	>8	

5f	8	>1.6	>1.6	>1.6	>1.6	>1.6	200	>40	>40	>40	40	>8	>8	>8	>8	>8
Brivudin(μ M)	>250	0.08	10	2	>250	>250	>250	>250	>250	>250	>250	>250	>25	>25	>250	>250
Ribavirin(μ M)	>250	250	250	150	150	>250	>250	30	150	50	>250	150	150	>25	>250	25
Acyclovir(μ M)	>250	0.4	0.4	>25	>250	50							0	0		0
Ganciclovir(μ M)	>100	0.032	0.006	100	>100	2.4							4			
(S)-DHPA(μ M)							>250	150	150	>250	>250	50	250	>25	>250	>250
													0	0		0

TABLE 4. Cytotoxicity and antiviral activity of compounds 4a-g and 5a-f against cytomegalovirus (CMV) and varicella-zoster virus (VZV) in human embryonic lung (HEL) cells.

Compd.	Antiviral activity EC ₅₀ (μ g/ml) ^a				Cytotoxicity (μ g/ml)		
	CMV (cytomegalovirus)		VZV (varicella-zoster virus)		Cell morphology MCC ^b		Cell growth CC ₅₀ ^c
	AD-169 strain	Davis strain	TK ⁺ (OKA strain)	TK (07/1 strain)	CMV assay	VZV assay	
4a	>4	>4	>4	>4	20	20	10.5
4b	>20	>20	>20	>4	100	≥20	>100
4c	>4	>4	>4	>4	20	20	12.6
4d	>20	>4	>4	>20	≥20	≥20	>100
4e	>20	>20	>20	>20	100	≥20	>100
4f	>20	>100	>20	>20	≥100	100	>100
4g	>20	>20	>20	>20	100	100	>100
5a	>20	>4	>4	>20	≥20	≥20	>100
5b	>20	>20	>0.8	>4	100	≥0.8	>100
5c	>4	>4	>4	>4	20	≥4	62.2
5d	>4	>20	>4	>4	≥20	20	>100
5e	>20	>20	>4	>4	100	20	>100
5f	>20	>4	>4	>4	≥20	≥4	>100
Ganciclovir	1.4	1.7	-	-	400	-	80
Cidofovir	0.24	0.37	-	-	400	-	57
Acyclovir	-	-	1.0	15	-	>50	190
Brivudin	-	-	0.0095	12.6	-	>50	244

^aEffective concentration required to reduce virus-induced cytopathic effect by 50%. Virus input was 20 (VZV) or 100 (CMV) plaque forming units (PFU).

^bMinimum cytotoxic concentration that causes a microscopically detectable alteration of cell morphology.

^cCytotoxic concentration required to reduce cell growth by 50%.

HEL cell culture : Herpes simplex virus type 1 (HSV-1) (KOS strain), Herpes simplex virus type 2 (HSV-2) (G strain), Vaccinia virus , Vesicular stomatitis virus , HSV-1 thymidine kinase deficient virus (TK-KOS ACV^r).

(c) HEL cell culture: Cytomegalovirus (CMV) (strains AD-169 and Davis), Varicella-zoster virus (VZV) (TK+VZV strain OKA strain and 07/1 strain).

(d) MT-4 cells: HIV-1 (IIIB) and HIV-2 (ROD) strains.

Brivudin, (S)-DHPA, ribavirin, acyclovir, cidofovir and ganci-clovir were used as the reference compounds. In the tests with viruses described in (a), (b), (c), (d) and (e) antiviral activity and cytotoxicity were determined with the compounds 4a-g and 5a-f. None of synthesized compounds had selective activity at subtoxic concentrations against the viruses tested.

Antitubercular activity

Compounds 4a-g and 5a-f were also tested for in vitro antitubercular activity against *M. tuberculosis* H37Rv (ATCC 27294) using the BACTEC 12B medium and a broth microdilution assay, the Microplate Alamar Blue Assay (MABA) (28,29). Rifampicin was used as the standard in the antitubercular assays. None of the tested compounds were considered for further antitubercular evaluation as they exhibited less than 90% inhibition in the primary screen (MIC > 6.25 μ g/mL).

Anticancer activity

Both cytotoxicity and anticancer assay results showed that, none of the tested concentrations of the compounds gave IC_{50} values. Therefore, it was concluded that there were no significant differences found between cytotoxic and anticancer ref- caused 10-20% cytotoxic effect at the highest concentration on 4th day of the incubation period (30) (Figure 2)

EXPERIMENTAL

Chemistry

All chemical compounds were purchased from Fluka. Melting points were taken on Buchi-530 apparatus. Merck silica gel 60 F254 plates were used for analytical TLC and visualized with UV. The IR spectra were obtained with a Shimadzu FTIR-8400. ¹H NMR spectra in DMSO-*d*₆ were obtained on a Bruker Avance-DPX 400 instrument. HR-Mass spectra using EI and FAB ionization techniques, were performed using a Jeol JMS-700 instrument.

Synthesis of 4-Aminobenzoic acid hydrazide 1 (17)

Ethyl 4-aminobenzoate (0.01 mol) was added to hydrazine-hydrate (99%, 3 mL). The reaction mixture was heated for 1 h and this reaction mixture was refluxed in the presence of etha-nol. The compound thus obtained was allowed to stand over-night. The precipitated solid was washed with water, dried and cleaned twice using hot methanol.

4-Amino-N'-[(4-fluoro/4-(trifluoromethyl)phenyl)methylene] benzohydrazide 2 (17), 3

General procedure

A solution of 0.01 mol of **1** and equimolar amount of appropriate aldehyde in 60 mL of ethanol was heated under reflux for 1 h (15 min for compound **3**). The precipitate obtained was filtered off, washed with water and cleaned twice with boiling EtOH.

zone);

TABLE 5. Cytotoxicity and antiviral activity of compounds 4a-g and 5a-f against HIV-I (IIIB) and HIV-II (ROD).

Compounds	HIV-I(IIIB)		HIV-II(ROD)	
	EC ₅₀ (μ g/ml) ^a	CC ₅₀ (μ g/ml) ^b	EC ₅₀ (μ g/ml) ^a	CC ₅₀ (μ g/ml) ^b
4a	>53.85	53.85	>53.85	53.85
4b	>125	>125	>125	>125
4c	>20.3	94.00	>16.8	94.00
4d	>125.00	>125.00	>125.00	>125.00
4e	>125.00	>125.00	>125.00	>125.00
4f	>59.10	59.10	>59.10	59.10
4g	>125.00	>125.00	>125.00	>125.00
5a	>70.9	>125	>79.2	>125
5b	>125.00	>125.00	75.25	>125.00
5c	>58.10	>58.10	>58.10	>58.10
5d	>68.10	68.10	>68.10	68.10
5e	>75.70	75.70	>75.70	75.70
5f	>118.00	>125.00	>125.00	>125.00

^aEffective concentration required to protect 50% of the cells against destruction by the virus.

^bCytotoxic concentration required to destroy 50% of the uninfected host cells.

(d, 2H, o-NH₂, J=8.6Hz), 7.69(d, 2H, m-NH₂, J=8.6Hz), 7.80(d, 2H, o-CH, J=8.3Hz), 7.91(d, 2H, m-NH, J=8.3Hz), 8.46(d, 1H, -CH=N), 11.65(s, 1H, -CONHN=CH-); HR-MS(EI, 70eV): m/z (calculated/found) for C₁₅H₁₂F₃N₃O₃ 307.0932[M⁺], 307.0903.

1-[4-[[2-[(4-substituted phenyl)methylene]hydrazino]carbonyl]phenyl]-3-substituted thioureas 4a-g, 5a-f

General procedure
A dry acetonitrile solution of 4-amino-N'-[(4-fluoro/4-trifluorophenyl)methylene]-benzohydrazide and equimolar substituted phenyl isothiocyanates in dry acetonitrile was heated under reflux for 9-

15h. The completion of reaction was checked by TLC (petroleum ether: acetone, 50:50, v/v). The precipitate obtained was filtered off and recrystallized twice with dry acetonitrile.

Biological activity

Antiviral activity

Compounds **4a-g** and **5a-f** were tested for antiviral activity and cytotoxicity in various viral test systems, according to previously published procedures (23-27). The synthesized compounds were tested against HIV-1 (IIIB) and HIV-2 (ROD), vesicular stomatitis virus, Coxsackie B4 virus, respiratory syncytial virus, parainfluenza-3 virus, reovirus, Sindbis virus, Punto Toro virus, herpes simplex virus type 1 and 2 and vaccinia virus-induced cytopathogenicity at subtoxic concentrations in MT-4 cells, HeLa, Vero or He1 cell culture. Brivudin, (S)-DHPA, ribavirin, acyclovir, cidofovir and ganciclovir were used as the reference compounds.

Antitubercular activity

Antitubercular evaluation was carried out in the Tuberculosis

Antimicrobial Acquisition and Coordinating Facility (TAACF).

Primary screen was conducted at 6.25 µg/ml against *M. tuberculosis* H37Rv in BACTEC 12B medium using both BACTEC 460 radiometric system and Microplate Alamar Blue Assay (MABA) (28, 29). Compounds effecting < 90 % inhibition in the primary screen (MIC > 6.25 µg/ml) were not further evaluated. Compounds demonstrating at least 90 % inhibition in the primary screen were considered for re-testing at lower concentration (MIC) in a broth microdilution MABA.

Anticancer activity

The synthesized compounds were tested for anticancer activity and cytotoxicity. The Cell Titer 96 Aqueous ONE Solution (Promega, Madison, WI) was used to evaluate cellular viability utilizing reduction of 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (MTS).

Cell culture and viability assay

To assess cytotoxicity and anticancer effects, the A 549 and L 929 cell lines were used. The cells were cultured in a 75 mm flask with 5% CO₂ and passed every three days. We used the MTS test to check the cell viability. The cells were cultured in a 75 mm flask with 5% CO₂ and cultured every three days according to protocol. We used the MTS test to check the cell viability. A 96-well tissue culture plate was used, with 5,000 cells per well. The cells were cultured for 4 days (30) after the media was changed to include different concentrations of chemicals (10 nM, 100 nM, 1 µM, and 10 µM).

The MTS assay was carried out in accordance with the manufacturer-provided protocol. Briefly, 20 microliters of MTS solution was added to each well, and the cells were incubated at 37°C for 1 to 3 hours. Next, we measured the wells' absorbance at 490 nm. The data was expressed as a percentage of the values obtained from chemical-free cell cultures grown under identical circumstances. The L929 cells were exposed to the same dose of chemicals used to detect the carcinogen effect in order to conduct the time-course study of their cytotoxicity. The viability of the cells was assessed using the MTS assay one to four days after treatment began.

A final dosage of 0.1% DMSO was used to dissolve all test substances. In these tests, the solvent failed to exhibit any activity at the screening level. Doxorubicin and taxol were chosen as reference medicines for the purpose of comparing the test compounds' anticancer activity and cytotoxicity testing.

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